## **Advances in Crop Science and Technology**

**Short Communication** 

**Open Access** 

Received April 15, 2015; Accepted June 13, 2015;

<sup>\*</sup>Corresponding author: Mariam Sticklen, Department of Plant, Soil and Microbial Sciences, Michigan State University, East Lansing, MI 48824, USA, Tel: 1 517-355-0271; E-mail: stickle1@msu.edu

with standardized appropriate parts, and with predictable and reliable expected functions [9].

Because the synthesis of most metabolic pathways such as the one for oil biosynthesis or for plant cell wall biosynthesis circuits are so large, scientists working on such synthetic biology also need to use the routine engineering techniques called "decoupling and abstraction" i.e. breaking of the whole circuits into smaller modules or pieces, and testing of each pieceprior to their assembly [2].

Like in engineering, the prefabricated parts such as DNA pieces must be combined following the engineering rules of "assembly standard" including speed, versatility, laboratory autonomy and full combinational potential where the parts can become interchangeable. Despite the above rules, the need for mathematical modeling of each piece of the circuits or each prefabricated part is important in order to assure the rationality of the designed genetic circuits [2].

Also, in an ideal 21<sup>st</sup> century a ordable manufacturing scenario, the pre-fabricated parts of any circuits should also be potentially used in di erent synthetic biology hosts such as di erent crops.

A very powerful DNA assembly system called "Golden Braid" has been used in plant synthetic biology research. is DNA assembly system contains DNA building modules (parts) to be used in di erent crop synthetic biology approaches, a system that can be optimized for many uses. e Golden Braid DNA assembly modules arealready commercially sold in form of a "tool kit" for their uses in plant synthetic biology research [1].

e authors team used the techniques of RNAi genetic engineering and produced a corn crop that its cell walls contains about 8% less lignin, and therefore not only its stover could be converted into fermentable sugars with less needs for pretreatment processes, but also the energy saved by plants by producing less lignin was shi ed to producing more cellulose resulting in more fermentable sugars [10,11] Using synthetic biology of corn crop, it might be possible to improve the systen by rst logically designing the circuits network, measuring the stability of its steady-state, modelling their behaviors, and nally assembling the system in form of genetic circuits with standardized appropriate parts, and with predictable and reliable expected functions.

erefore the crop synthetic biology is much more complicated, but also more precise with better predictable results.

e author's team also produced a corn crop that its stover expresses all three microbial cellulase enzymes (endoglucanse, exoglucanase and beta-glucosidase) needed to convert the plant cellulose into fermentable sugars. is transgenic corn cropself-produces all of these three cellulases in its stover (not in owers, seeds or roots) and such stover could be converted into fermentable sugars without the needs for any externally applied microbial cellulases [12]. If the system was to be synthetically made using the technologies associated with synthetic biology, it should be possible to logically design circuits networks of crop cell walls consisting of cellulase genes, measure the stability of their steady-state, model their behaviors, and assemble the system in form of genetic circuits with standardized appropriate parts, and apply the system for production of a novel corn crop with predictable and reliable expected functions of self producing cellulases.

It might be also possible to combine both of the above examples to produce a bio energy crop with ideal cell walls that have high cellulose and hemicelluloses molecules; but with stronger but yet more bioprocessable lignin contents.

Plant synthetic biology can also include multiple genetic regulatory

system designs [13], such as switches or promoters to turn on and o the genes as they are required [2].

Synthetic biology of *E.* -producing butanol by Michelle Wang's team is a great example of the potential of the powerful synthetic biology [14]. It is also encouraging to see the most recent advances in comprehensive DNA assembly framework developed for plant synthetic biology [1].

In using the plant synthetic biology, it is also possible to predict the directed evolution and use such predictions to design novelcrops similar to those selected in vivo for the best traits [15], or produce novel cereal crops that can x nitrogen, x carbon more e ciently with a much more e ective photosynthesis, produce and store more sugars, have water use e ciency, disease and pest resistance and more nutritious seeds and vegetative tissues.

Despite all of its prmisses that potentially can advance the foold and energy security and produce low-coss industrial products, there are multiple challenges associated with the application of the crop synthetic biology. e challenges are; (1) unavailability of most naturally existing biological genetic pathway systems and the ambiguity of the certain genes involved such as those in cell wall synthesis pathway [16] or plant fatty acid biosynthesis and assembly pathways for oil production [17], (2) limited and impeding ability to rationally designthelanguage of the plants genetic system [18,19] and its gene circuits for the nest predictable direct evolution, (3) the unpredictable evolution of the genes' regulatory sequences, and (4) the acceptance of synthetically produced crops by the public.

While more fundamental research is needed to reduce most of the above challenges, the author expects major public concerns on the applications of synthetic biology. However, such concerns would lessen as soon as the bene ts of synthetic biology become more obvious to the public. Public needs to become educated by social scientists and by the media on the most humanitarian bene ts of the system biology, such as the one developed for the creation of the award winning anti-malarial drug produced through synthetic microbes [20]. Such drug can save millions of human lives in the developing nation where malaria is the most cause death, especially in children.

## References

- Sarrrion-Perdigones M, Vazquez-Vilar J, Palaci B, Castelijns J, Forment, et al. (2013) GoldenBraid 2.0: A comprehensive DNA assembly framework for plant synthetic biology. Plant Physiol 162: 1618-1631.
- Bowen TA, Zdunek JK, Medford JI (2008) Cultivating plant synthetic biology from system biology. New Phytologist 179: 583-587.
- 3. Drublin DA (2007) Designing biological systems. Genes & Development 21: 242-254.
- Elowitz MB, Leibler S (2000) A synthetic oscillary network of transcriptional regulators. Nature 403: 335-338.
- Basu S, Gerchman Y, Collins CH, Arnold FH, Weiss R (2005) A systemic multi cellular system for programmed pattern formation. Nature 434: 1130-1134.
- Andrianov V, Borisjuk N, Pogrebnyak N, Brinker A, Dixon J, et al. (2010) Tobacco as a production platform for biofuel: Overexpression of Arabidopsis DGAT and LEC2 genes increases accumulation and shifts the composition of lipids in green biomass. Plant Biotechnol. J 8: 277-287.
- Slocombe SP, Cornah J, Pinfeld-Wells H, Soady K, Zhang Q, et al. (2009) Oil accumulation in leaves directed by modifcation of fatty acid breakdown and lipid synthesis pathways. Plant Biotechnol J 7: 694-703.
- Sticklen M (2013) An Advance in Crop Science and Technology: Potentials for Producing the High-Value High-Calorie Triacyglycerols Commodity in Crop Vegetative Wastes. Advances in Crop Sci. and Technology e104.

- 9. Andriantoandro E, Basu S, Kraig DK, Weiss R (2006) Synthetic biology: New engineering rules for an emerging discipline. Mol Sys Biol 2: 0028.
- 10. Park S-H, Mei C, Pauly RM. Garlock BE, Dale R. Sabzikar H (2012) Downregulation of Maize Cinnamoyl-CoA Reductase via RNAi Technology Causes Brown Midrib and Improves AFEX<sup>™</sup>-Pretreated Conversion into Fermentable Sugars for Biofuels. Crop Sci. 52: 2687-2701.
- Park S-H, Ong RG, Mei C, Sticklen M (2014) LigninDown-regulation of Zea mays via dsRNAi and klason lignin analysis. 89.
- 12. Park S-H, Ransom C, Mei C, Sabzikar R, Qi C, et al. (2011) In the quest of alternatives to microbial cellulase mix production: Corn stover-produced heterologous multi-cellulases readily deconstruct lignocellulosic biomass to fermentable sugars. J Chem Techno Biotechnol.
- Gardner TS, Cantor CR, Collins JJ (2000) Construction of a genetic toggle switch in Escherichia coli. Nature 403: 339-342.
- 14. Wen M, Bond-Watts BB, Chang CY (2013) Production of advanced biofuels in engineered Current Opinion in Chemical Biol 17: 472-479.

- Yokobayashi Y, Weiss R, Arnold FH (2002) Directed evolution of a genetic circuit. Proc Natl Acad Sci, USA. 99: 16587-16591.
- 16. Sticklen M (2007) Feedstock genetic engineering for biofules. Crop Sci 47: 2238-2248.
- Vanhercke T, ElTahchy A, Liu Q, X-Rong Zhou, Shrestha P, et al. (2014) Metabolic engineering of biomass for high energy density: oilseed-like triacylglycerol yields from plant leaves. Plant Biotechnol J 12: 231-239.
- Pedersen M, Phillips A (2009) Towards programming languages for genetic engineering of living cells. J Royal Society of Interface.
- Umesh P, Naveen F, Rao MA, Nair S (2010) Programming languages for synthetic biology. Syst Synth Biol 4: 265-269.
- Priscilla EM, Purnickand R, Weiss (2009) The second wave of synthetic biology: from modules to systems. Nature Reviews Mol Cell Biol 10: 410-422.

**Citation:** Sticklen M (2015) Advances in Crop Synthetic Biology: A Platform that Bridges the Expertise of Crop Molecular Biologists, Engineers, Software Developers and Mathematical Modelers. Adv Crop Sci Tech 3: 171. doi:10.4172/2329-8863.1000171