

Biosensors for Indirect Monitoring of Foodborne Bacteria

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Abstract

Microbiological safety assessment in food samples is often a cumbersome and time consuming task that has to be done in a regular basis in most food manufacturing facilities. Standard microbiological techniques for evaluating critical levels of bacteria take several days to yield results, thus the food products have to be withheld in the processing associated with bacterial activity are amongst the most promising technologies for addressing this need. State-of-the-art biosensors capable of real-time monitoring of biomarkers from foodborne bacteria.

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Villalonga et al. demonstrated a biosensor enhanced with single-walled carbon nanotubes (SWNT) and gold nanoparticles (AuNP) for the direct determination of xanthine, which is a common marker of bacterial spoilage in meat. The operating mechanism of this electrochemical device is based on an enzymatic reaction catalyzed by xanthine oxidase (XO), immobilized on the surface of a pAuNP/SWNT/glassy carbon electrode (GCE) [9]. Ko et al. presented a biosensor consisting of a chip functionalized with gold nano-islands, single-walled carbon nanotubes (SWNT) and a 25 mer p-DNA for selective detection of t-DNA. The sensing principle is based on changes in the overall conductance occurring after hybridization of t-DNA with p-DNA on the chip surface [10]. Lata et al. developed a D-amino acid biosensor based on covalent immobilization of D-amino acid oxidase (DAAO) onto a gold electrode (Au) modified with multi-walled carbon nanotubes (c-MWCNT), copper nanoparticles (CuNPs) and polyaniline (PANI). In this study, the authors only aimed to quantify the level of amino acids in fruit juices as a measure of their nutritional value; nonetheless, the device could also be used to estimate presence and maybe even viability of bacteria based on the relative amounts of D and L amino acids and the rate of racemization [11]. Wang et al. created a biomimetic sensor based on graphene, gold nanoparticles (Gr-AuNPs) and molecularly imprinted polymers (MIP) onto a glassy carbon electrode (GCE) for detecting glycoproteins. In this case, a BSA template was used as model protein for technology demonstration. An amperometric device could be applied for detecting cell membrane glycoproteins which are known to be important virulence factors in pathogenic microorganisms such as *E. coli* and *Streptococcus* spp. [12]. Cao et al. built an electrochemical biosensor functionalized with graphene, platinum, palladium, chitosan (GS-CS-PtPd) and cholesterol oxidase (ChOx). The probe was designed for cholesterol sensing in food samples, and has a potential use for monitoring the transformation of sterols by specific spoilage microorganisms [13].

Challenges and opportunities

Enzymatic biosensors for monitoring biomarkers are highly

agents, hydrogels based on weakly ionizable polysaccharides show pH-responsive phase transition [23]. is stimulus-response behavior can be exploited in biosensors for controlled release of small molecules and scaffolding of enzymes.

Conclusions

The long time required for testing the microbiological safety and quality of food products using standard microbiological methods is a concerning problem for the food industry and public health. Biosensor technologies that target bacterial biomarkers are promising alternatives for rapid screening of harmful bacteria in food samples. Incorporation of nanomaterials with unique electrical and photonic properties, as well as biomaterials with high biocompatibility are the most effective strategies for developing biosensors with ultrafast response time and high stability.

References

1. Centers for Disease Control and Prevention. (2011) *Foodborne Illness in the United States*. Retrieved February 25, 2016, from <http://www.cdc.gov/foodborneburden/2011-foodborne-estimates.html>
2. U.S. Food and Drug Administration. (2015). *Food: Compliance and Enforcement*. Retrieved February 25, 2016, from <http://www.fda.gov/Food/ComplianceEnforcement/default.htm>
3. Denoya C, Colgan S, du Moulin GC. Alternative Microbiological Methods in the Pharmaceutical Industry: The Need for a New Microbiology Curriculum. *Am. Pharm. Rev* 9: 10-18.
4. Zhao X, Lin CW, Wang J, Oh DH (2014) Advances in Rapid Detection Methods for Foodborne Pathogens. *J Microbiol Biotechnol* 24: 297-312.
5. Vanegas DC, Gomes C, McLamore ES (2014) Xanthine Oxidase Biosensor for Monitoring Meat Spoilage. *Proc. SPIE* 9107.
6. Compagnone D, Trojanowicz M (2007) Enantioselective screen-printed amperometric biosensor for the determination of D-amino acids. *Bioelectrochemistry* 71: 91-98.
7. Henao-Escobar W, del Torno-de Román L, Domínguez-Renedo O, Alonso-Lomillo MA, Arcos-Martínez MJ (2016) Dual enzymatic biosensor for simultaneous amperometric determination of histamine and putrescine. *Food Chem* 190: 818-823.
8. Ferracci G, Marconi S, Mazuet C, Jover E, Blanchard MP, et al. (2011) A label-free biosensor assay for botulinum neurotoxin B in food and human serum. *Anal Biochem* 410: 281-288.
9. Villalonga R, Díez P, Eguílaz M, Martínez P, Pingarrón JM (2012) Supramolecular immobilization of xanthine oxidase on electropolymerized

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