

Further, on adding graphene sheets to the prepared bioprobe the fluorescence was recorded as switch on and after addition of various concentrations of cTnI, again the fluorescence recovered in photoluminescence spectra. The sensor takes only 10 min with low detection limit of 0.192 pg/mL. Similarly, Shankar et al., proposed immobilized a 16-phosphonohexadecanoic acid self-assembled monolayer (SAM) onto a TiO₂ array to constitute a low-cost biosensor [44]. On this ultrasensitive platform, the detection of cTnI concentrations as low as 0.1 pg mL⁻¹ was accomplished with the help of enzymatic amplification. The reported study takes over 2 h to complete the analysis. Similarly, another way to design a biosensor when a fluorophore is close to the metal surface, its optical properties will significantly alter. Based on this metal quenching effect, the detection sensitivity can be improved. Lee and Kang deposited gold onto the glass substrate, followed by immobilizing protein A, anti-cTnI, cTnI antigen, and fluorescence labeled anti-cTnI, respectively [45]. Depending on the distance between the dye and the gold surface, the fluorescent dye was not greatly quenched, but adsorption of dye led to the quenching of its fluorescence. Therefore, such enhanced fluorescence was detectable about 7000 times lower in the detection limit compared to the traditional method. However, it is still a time-consuming procedure. Alkaline phosphatase (ALP) has been largely exploited for the fluorescent based cTnI immunoassay, especially in commercial products and recently has explored the ALP chemiluminescence chemistry for the cTnI detection [25,46-48]. The combination of magnetic and fluorescence strategy is promising for the quick and sensitive for cTnI detection. In both the cases, the investigation time is about 40 min which is better in comparison to the TiO₂ nano-array and the limit of detection for both can attain as low as 1 ng/mL.

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