Vjg"Dwtfgp"qh"JKX"Cuuqekcvgf"Ftwi"Tgukuvcpeg"Owvcvkqpu"kp"cp"Gctn{"Kphcpv Fkcipquku"Rtqitco<"C"Incpeg"vjtqwij"vjg"Rcgfkcvtke"Ykpfqy"qh"\kodcdyg

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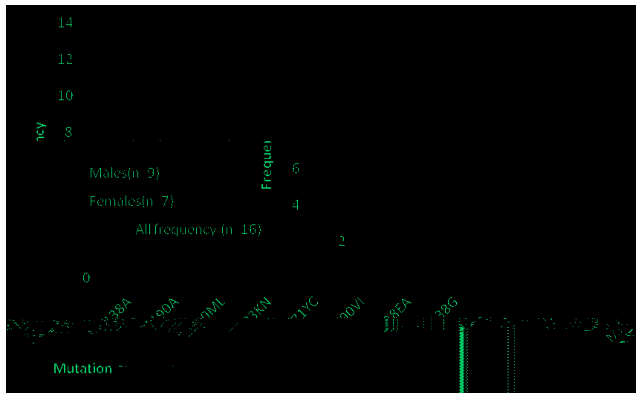
cycling conditions consisted of a denaturation of 94C for 5minutes followed by 35 cycles of 94C for 30 seconds, 55C for 30 seconds, an extension of 72C for 1minute and a final extension at 72C for 7 minutes. The cycling conditions for the second round PCR were similar to those for the first round PCR. After the second round PCR, a 684 base pair DNA fragment, corresponding to the amplified HIV-1 C product was assessed by ethidium bromide stained 1% agarose gel electrophoresis. A positive control containing 10 copies of HIV-1 C was included in each PCR reaction. Negative controls containing all the PCR reagents excluding template DNA were also included in every PCR run to ensure there was no contamination of any of the reagents.

HIV reverse transcriptase sequencing

Samples that gave the characteristic 684bp band were sequenced by the dye terminator method as described [33], briefly; PCR amplicons were cleaned up using the PureLink QUICK PCR Purification Kit (Life



14
12
10
8
6
4
2
0
Males(n 9)
Females(n 7)
All frequency (n 16)
Mutation

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