

The Sensory Impact of Nicotine on Noradrenergic and Dopaminergic Neurons of the Nicotine Reward - Addiction Neurocircuitry

Jed E Rose¹, Ozra Dehkordi^{2,3*}, Kebreten F Manaye³, Richard M Millis⁴, Salman Ameri Cianaki² and Annapurni Jayam-Trouth²

¹Department of Psychiatry, Duke University Medical Centre, Durham, N.C. 27705, United States

²Department of Neurology, Howard University Hospital, Washington, D.C. 20060, United States

³Department of Physiology & Biophysics, Howard University College of Medicine, Washington, D.C. 20059, United States

⁴Department of Medical Physiology, American University of Antigua College of Medicine, St. John's, Antigua and Barbuda

*Corresponding author: Dehkordi Ozra, Associate Professor, Department of Neurology, Howard University Hospital, Washington, United States, Tel: +1 202 865 1978; Fax: +1 202 865 1977; E-mail: odehykordi@howard.edu

Received date: Feb 16, 2016; Accepted date: April 01, 2016; Published date: April 07, 2016

Copyright: © 2016 Rose JE, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

The sensory experience of smoking is a key component of nicotine addiction known to result, in part, from stimulation of nicotinic acetylcholine receptors (nAChRs) at peripheral sensory nerve endings. Such stimulation of nAChRs is followed by activation of neurons at multiple sites in the mesocorticolimbic reward pathways. However, the neurochemical profiles of CNS cells that mediate the peripheral sensory impact of nicotine remain unknown. In the present study in mice, we first used c-Fos immunohistochemistry to identify CNS cells stimulated by nicotine (NIC, 40 µg/kg, IP) and by a peripherally-acting analog of nicotine, nicotine pyrrolidine methiodide (NIC-PM, 30 µg/kg, IP). Sequential double-labelling was then performed to determine whether noradrenergic and dopaminergic neurons of the nicotine reward-addiction circuitry were primary targets of NIC and NIC-PM. Double-labelling of NIC and/or NIC-PM activated c-Fos immunoreactive cells with tyrosine hydroxylase (TH) showed no apparent c-Fos expression by the dopaminergic cells of the ventral tegmental area (VTA). With the exception of sparse numbers of TH immunoreactive D11 cells, dopamine-containing neurons in other areas of the reward-addiction circuitry, namely periaqueductal gray, and dorsal raphe, were also devoid of c-Fos immunoreactivity. Noradrenergic neurons of locus coeruleus (LC), known to innervate VTA, were activated by both NIC and NIC-PM. These results demonstrate that noradrenergic neurons of LC are among the first structures that are stimulated by single acute IP injection of NIC and NIC-PM. Dopaminergic neurons of VTA and other CNS sites, did not respond to acute IP administration of NIC or NIC-PM by induction of c-Fos.

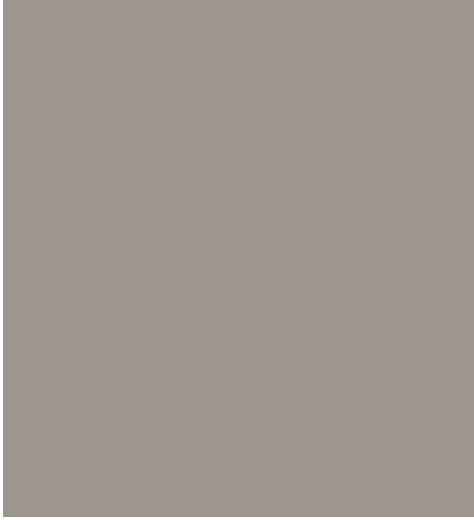


Figure 1: Fluorescent and laser scanning confocal microscopy images of representative brainstem sections demonstrating nicotine (NIC) and nicotine pyrrolidine methiodide (NIC-PM) activation of noradrenergic neurons of locus coeruleus (LC). Panels A-C: Control data demonstrating the effects of acute intraperitoneal injection of physiological saline (PS) on c-Fos activation of tyrosine hydroxylase (TH)-immunoreactive (IR) cells of LC. Panels D-I: Low power fluorescent (D-F) and high power confocal (G-I) images showing NIC-induced c-Fos IR cells (D, G), TH IR cells (E, H) and merge images of c-Fos with TH IR cells in LC. Panels J-L: High power confocal images showing NIC-PM induced c-Fos IR cells (J), TH-IR cells (K) and merge images of c-Fos with TH-IR cells (L) in LC. Arrows point to representative double-labeled neurons

Semi-quantification

In the most caudal extent of VTA (bregma -3.87 mm to -3.51 mm), NIC and NIC-PM activated c-Fos IR cells were sparsely scattered among the dopaminergic cells of paranigral nucleus (PN), parainterfascicular nucleus (PIF) and parabrachial pigmented nucleus (PBP). c-Fos IR cells were also detected at sites medial and ventral to the dopaminergic cells in regions which correspond to interpeduncular nucleus rostral (IPR) and pontine nuclei (Pr). More rostrally in the anterior extension of VTA (bregma -3.15 mm to -3.07 mm), NIC and NIC-PM activated cells were found mainly ventral and medial to the dopaminergic cells of VTA rostral (VTAR) and PBP, at sites which overlapped interfascicular nucleus (IF), rostral linear nucleus (RLi), IPR and retromamillary nucleus (RM) (Figure 3).

c-Fos IR cells were also seen medial and dorsal to the dopaminergic

analysis. Study concept and design: OD, JER. Acquisition of data: OD, SAC. Analysis and interpretation of data: OD, RMM. Drafting of the manuscript: OD, RMM. Critical revision of the manuscript for important intellectual content: OD, JER, RMM. Statistical analysis: OD, RMM. Obtained funding: OD, JER. Administrative, technical, and material support: KFM, A.JT. Study supervision: OD.

References

1. Wonnacott S, Barik J, Dickinson J, Jones IW (2006) Nicotinic receptors modulate transmitter cross talk in the CNS: nicotinic modulation of transmitters. *J Mol Neurosci* 30: 137-140
2. Dani JA, Bertrand D (2007) Nicotinic acetylcholine receptors and nicotinic cholinergic mechanisms of the central nervous system. *Annu Rev Pharmacol Toxicol* 47: 699-729
3. Albuquerque EX, Pereira EF, Alkondon M, Rogers SW (2009) Mammalian nicotinic acetylcholine receptors: from structure to function. *Physiol Rev* 89: 73-120
4. Zoli M, Pistillo F, Gotti C (2015) Diversity of native nicotinic receptor subtypes in mammalian brain. *Neuropharmacology* 96: 302-311.
5. Schultz W, Dayan P, Montague PR (1997) A neural substrate of prediction and reward. *Science* 275: 1593-1599
6. Wise RA (2009) Roles for nigrostriatal—not just mesocorticolimbic—dopamine in reward and addiction. *Trends Neurosci* 32: 517-524
7. Schultz W (2010) Dopamine signals for reward value and risk: basic and recent data. *Behav Brain Funct* 6: 24
8. Ikemoto S (2010) Brain reward circuitry beyond the mesolimbic dopamine system: a neurobiological theory. *Neurosci Biobehav Rev* 35: 129-150
9. De Biasi M, Dani JA (2011) Reward, addiction, withdrawal to nicotine. *Annu Rev Neurosci* 34: 105-130
10. Gu Q, Ni D, Lee LY (2008) Expression of neuronal nicotinic acetylcholine receptors in rat vagal pulmonary sensory neurons. *Respir Physiol Neurobiol* 161: 87-91.
11. Rose JE, Zinser MC, Tashkin DP, Newcomb R, Ertle A (1984) Subjective response to cigarette smoking following airway anesthetization. *Addict Behav* 9: 211-215
12. Rose JE, Tashkin DP, Ertle A, Zinser MC, Lafer R (1985) Sensory blockade of smoking satisfaction. *Pharmacol Biochem Behav* 23: 289-293
13. Rose, JE, Westman EC, Behm FM, Johnson MP, Goldberg JS (1999) Blockade of smoking satisfaction using the peripheral nicotinic antagonist trimethaphan. *Pharmacol Biochem Behav* 62: 165-172
14. Alimohammadi H, Silver WL (2000) Evidence for nicotinic acetylcholine receptors on nasal trigeminal nerve endings of the rat. *Chem Senses* 25: 61-66
15. Dehkordi O, Rose JE, Balan KV, Kc P, Millis RM, et al. (2009) Neuroanatomical relationships of substance P-immunoreactive intrapulmonary C-fibers and nicotinic cholinergic receptors. *J Neurosci Res* 87: 1670-1678
16. Dehkordi O, Rose JE, Balan KV, Millis RM, Bhatti B, et al. (2010) Co-expression of nAChRs and molecules of the bitter taste transduction pathway by epithelial cells of intrapulmonary airways. *Life Sci* 86: 281-288
17. Dehkordi O, Rose JE, Asadi S, Manaye KF, Millis RM, et al. (2015) Neuroanatomical circuitry mediating the sensory impact of nicotine in the central nervous system. *J Neurosci Res* 93: 230-243
18. Corrigan WA, Coen KM (1991) Selective dopamine antagonists reduce nicotine self-administration. *Psychopharmacology (Berl)* 104: 171-176
19. Bromberg-Martin ES, Matsumoto M, Hikosaka O (2010) Dopamine in motivational control: rewarding, aversive, and alerting. *Neuron* 68: 815-834
20. Pistillo F, Clementi F, Zoli M, Gotti C (2015) Nicotinic, glutamatergic and dopaminergic synaptic transmission and plasticity in the mesocorticolimbic system: focus on nicotine effects. *Prog Neurobiol* 124: 1-27.
21. Li S, Shi Y, Kirouac GJ (2014) The hypothalamus and periaqueductal gray are the sources of dopamine fibers in the paraventricular nucleus of the thalamus in the rat. *Front Neuroanat* 8: 136
22. Gillis CN, Lewis JJ (1956) The pharmacology of nicotine monomethiodide. *J Pharm Pharmacol* 8: 46-54
23. Aceto MD, Awaya H, Martin BR, May EL (1983) Antinociceptive action of nicotine and its methiodide derivatives in mice and rats. *Br J Pharmacol* 79: 869-876
24. Lenoir M, Tang JS, Woods AS, Kiyatkin EA (2013) Rapid sensitization of physiological, neuronal, and locomotor effects of nicotine: critical role of peripheral drug actions. *J Neurosci* 33: 9937-9949
25. Cox BM, Goldstein A, Nelson WT (1984) Nicotine self-administration in rats. *Br J Pharmacol* 83: 49-55
26. Donny EC, Caggiula AR, Knopf S, Brown C (1995) Nicotine self-administration in rats. *Psychopharmacology (Berl)* 122: 390-394
27. Rose JE, Corrigan WA (1997) Nicotine self-administration in animals and humans: similarities and differences. *Psychopharmacology (Berl)* 130: 28-40 d

43. Flavin SA, Winder DG (2013) Noradrenergic control of the bed nucleus of the stria terminalis in stress and reward. *Neuropharmacology* 70: 324-330
44. Grenhof J, Nisell M, Ferré S, Aston-Jones G, Svensson TH (1993) Noradrenergic modulation of midbrain dopamine cell firing elicited by stimulation of the locus coeruleus in the rat. *J Neural Transm Gen Sect* 93: 11-25
45. Lategan AJ, Marien MR, Colpaert FC (1990) Effects of locus coeruleus lesions on the release of endogenous dopamine in the rat nucleus accumbens and caudate nucleus as determined by intracerebral microdialysis. *Brain Res* 523: 134-138

46.