

After cell treated with UNBS5162 for 24 h, cell migration and invasion were tested using the transwell assay in accordance with producer's instructions. For examining cell migration, 300 μ l serum-free medium containing with 1×10^5 cell was plated into the top of transwell chamber (Millipore Corp., Billerica, MA, USA) and 500 μ l medium with 10% FBS was plated into the low-storied chamber. After cultivation of 12 h, cells on the upper chamber were eliminated using cotton swabs and cells on the low-storied chamber were fixed in 4% paraformaldehyde, then stained with 0.1% crystal violet. Next, five random fields were chosen under microscope, photographed and counted. For examining cell invasion, Matrigel (BD Biosciences, San Diego, CA, USA) dissolved with serum-free DMEM diluted at 1:6 overnight in advance and paved to the upper chamber of transwell, the remaining steps resembled migration assay [17].

Cell proteins were extracted with RIPA lysis buffer and inhibitors specific to protease and phosphatase (Beyotime). Proteins concentrations were tested by bicinchoninic acid (BCA) (Beyotime) kit. Then, protein with equal amount (20 μ g) was separated in 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and transferred onto polyvinylidene difluoride (PVDF) membranes. After blocking with 5% BSA for 1.5 h, membranes were cultivated with primary antibodies at 4°C overnight and then incubated with secondary antibodies in dark place at room temperature for 2 h. An enhanced-chemiluminescence (ECL, Beyotime) detection system was applied to visualize the bands. The gray value was scanned by Quantity One (Bio-Rad). GAPDH was used as the internal control. Membranes were incubated with the antibodies as follows: primary antibody rabbit anti-human, AKT, p-AKT, mTOR,

p-mTOR, p-P70S6K, Bcl-2, Bax, Active Caspase-3 (at dilute of 1 : 1000, Cell Signaling Technology, Danvers, MA, USA), GAPDH (at dilute of 1:5000, Proteintech Group, Inc, Rosemont, IL, USA); corresponding horseradish peroxidase (HRP)-labelled secondary antibody (at dilute of 1:5000, Proteintech Group, Inc).

After 10 μ M UNBS5162 treating U251 cells for 24 h, cells apoptosis percentage was tested by Annexin V-FITC/Propidium Iodide (PI) and flow cytometry analyses in light of producer's manuals. After cell were collected, centrifuged and resuspended, 1×10^6 binding buffer were added to resuspend cells, and cells density were adjusted to 3×10^6 /ml. Cells were stained with 5 μ l Annexin V-FITC and 10 μ l PI (Beyotime) in dark place for 20 min. Finally, cells were detected and analyzed using a FACS Calibur instrument (BD Biosciences).

SPSS18.0 statistical analysis software (SPSS Inc., Chicago, IL) and GraphPad Prism 6.0 (GraphPad Software, Inc., LaJolla, CA, USA) were utilized to analyze the experimental data. The data are exhibited as the mean \pm standard deviation (SD). The Student's t test, ANOVA and post hoc Dunnett were used to analyze statistical correlation of data. $p < 0.05$ indicated statistically remarkable difference.

To assess the impact of UNBS5162 on GBM, we chose glioma cell U251 and the normal cell HEB using CCK-8 assay. 10 μ M and 100 μ M UNBS5162 treatment were effective on U251 cells (Figure 1A, $p < 0.01$). The viability of HEB cells was repressed only in 100 μ M UNBS5162

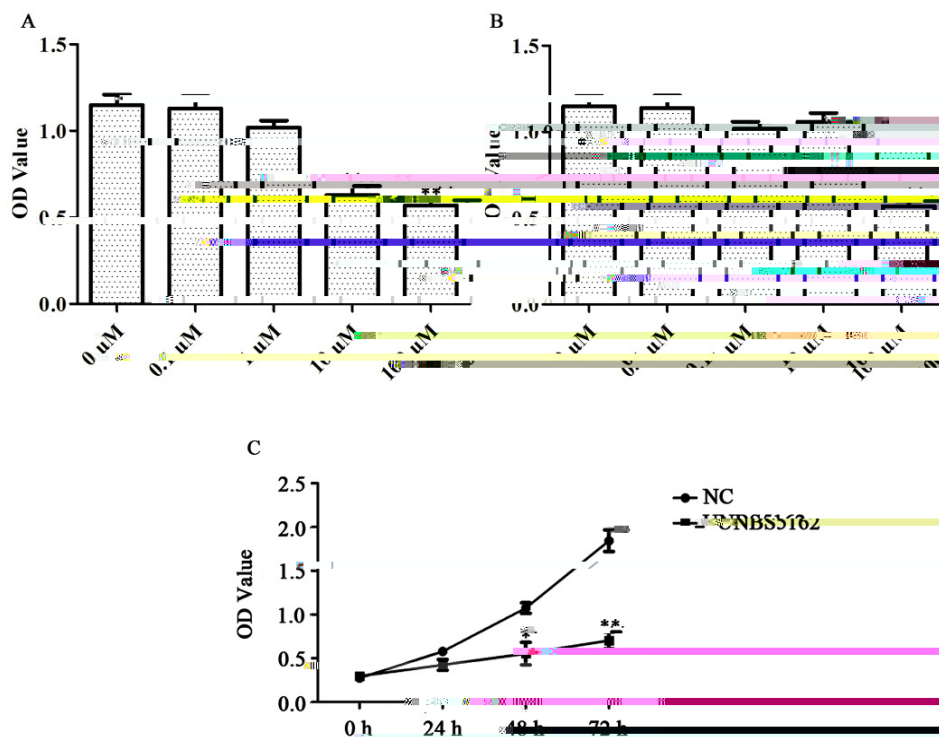


Figure 1: UNBS5162 inhibited U251 cell proliferation by the CCK-8 assay

treated group (Figure 1B, $p < 0.01$). So in the following experiments, 10 μM UNBS5162, a highly-effective and low-toxicity dose, was used. And the results also show that UNBS5162 obviously decreased the OD value of U251 cells in a time-dependent mode (Figure 1C, $p < 0.05$, $p < 0.01$).

These data indicated that UNBS5162 hampered glioma cell U251 viability effectively.

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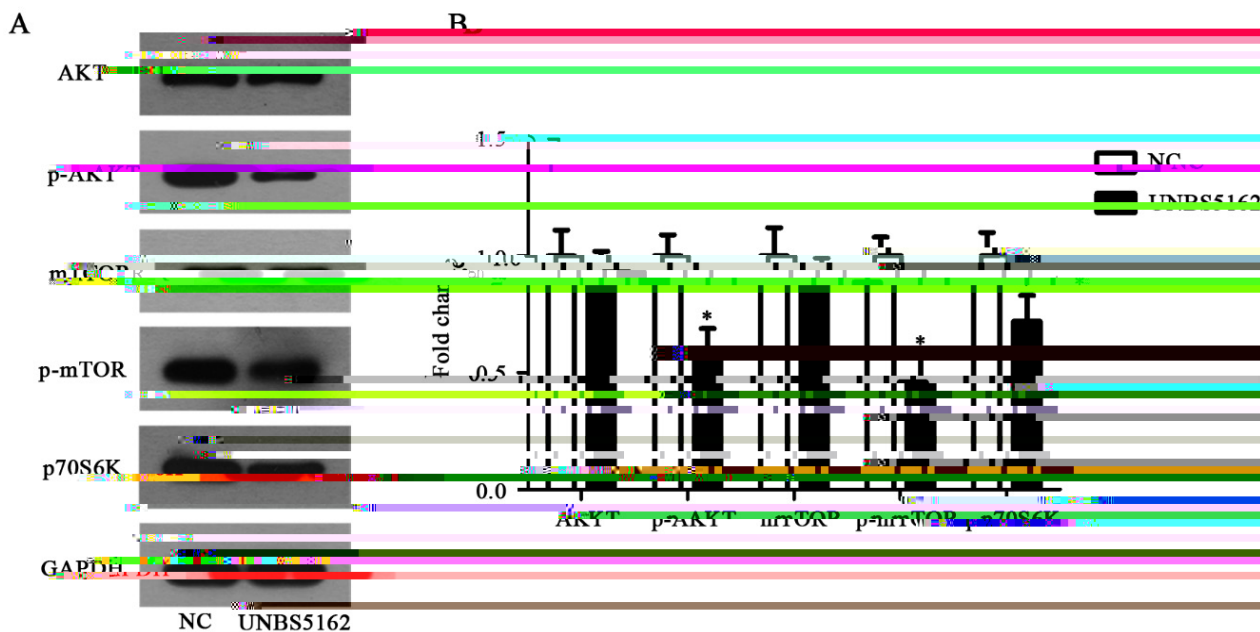


Figure 4: Western blot and bar graph showing protein levels of AKT, p-AKT, mTOR, p-mTOR, p70S6K, and GAPDH in U251 glioma cells treated with UNBS5162 compared to NC. The bar graph shows a significant decrease in p-AKT, p-mTOR, and p70S6K levels in the UNBS5162 group.



Figure 5: Bar graph showing the effect of different concentrations of UNBS5162 on U251 glioma cells viability. The graph shows a dose-dependent decrease in cell viability as the concentration of UNBS5162 increases.

esophageal, showing it has an effect on anti-angiogenesis. Lymphus, bone marrow, gastrointestinal tract and so on have been determined as targets for UNBS5162. Meanwhile, researches have suggested that *in vitro* incubation of UNBS5162 with human cancer cells obviously reduced the expression of the proangiogenic chemokines (e.g. CXCL1, CXCL2, CXCL3, CXCL5, CXCL6, and CXCL8). Because of these features, UNBS5162, as an antitumor agent, has considerable prospects to provide a new drug target for clinical cancer treatment and research. Researches have shown that UNBS5162 treatment can obviously reduce vascular surface area in human prostate cancer xenografts. But the functional action of UNBS5162 in other cancers remains unknown. In this work, we performed western blot assay, CCK8 proliferation test, transwell invasion/migration analyses and flow cytometry detection with U251 cell lines under different administration conditions. Our results suggested that UNBS5162 significantly repressed cell proliferation, migration/invasion capabilities and promoted cell apoptosis. In the future, UNBS5162 may become a potential molecular marker for therapy of glioma.

In our report, transwell assay showed that UNBS5162 inhibited glioma cell U251 invasion and migration. Furthermore, UNBS5162 could promote cell apoptosis and upregulate pro-apoptotic protein Active Caspase-3 and Bax, as well as downregulate anti-apoptotic protein Bcl-2. Bax/Bcl-2 and activation of Caspase-3, identified as a pivotal mechanism of action of anti-tumoral drug, are involved in cell death and apoptosis of tumor cells, which is developed as an important determining factor of chemotherapy effectiveness. Studies have shown that changes in tumor cell apoptosis are mainly due to abnormal expression of apoptosis-related factors and abnormal of cell apoptotic signal transduction pathways [21]. PI3K, known as a family of lipid kinases, plays an essential part in cell growth by inhibiting apoptosis in various cancers [22]. Our results showed that U251 cell apoptosis was associated with the PI3K pathway. In addition, PI3K pathway was involved in the development of cancer through AKT, mTOR and p-P70S6K, which suggests PI3K pathway to be the action pathways of UNBS5162 acting on cancer. The hallmarks p-AKT and p-mTOR in the pathway exert an important action in the proliferation and metastasis of tumor cells. Activation of Akt antagonizes angiogenesis, tumor metastasis and invasion and phosphorylates protein kinase mTOR [23]. Researchers have suggested that PI3K/AKT/mTOR pathway facilitates the resis
