

10 &)8 J ¿VK GD\ IRU

GD\ DQG PRQLWRUHG IRU VHUXP DQWLERG\ SURGXFWLRQ XS WR GD\ SRVW Y
 ¿VK UHFRUGHG WKH KLJKHVW DQWLERG\ WLWHU “ “ “
 DQG “ IROORZHG E\ WKDW IURP IUHH FHOO YDFFLQDWHG “ “ “
 “ “ “ DQG “ “ DQG FRQWURO ¿VK “ “ “
 “ “ “ DQG “ DW DQG GD\ SRVW YDFFL

Keywords: Bio Im vaccine; *Aeromonas hydrophila*; Enzyme Linked Immunosorbant Assay; *Labeo rohita*

Introduction

Culture of Indian major carps (IMC) contributes more than 80% of the total aquaculture production in India. *Labeo rohita* (rohu) with its high consumer preference and good growth rate, it is widely cultured in Indian subcontinent [1-3]. There has been a phenomenal shift from extensive to intensive culture of carps in the last three decades. Intensive aquaculture offers an increased opportunity for spreading of infectious diseases at all stages of production [4]. Among the bacterial pathogens *Aeromonas hydrophila* is a ubiquitous secondary pathogen of IMC including rohu. Several instances of infections with *A. hydrophila* in India have been reported in IMC in recent past [5] hence; vaccination of aquacultured fish is becoming inevitable with increasing health risks.

Since, the aim of vaccination is to increase the immune memory/ humoral antibody response against the specific antigen, the detection of these specific antibodies is very essential to evaluate the efficacy of a vaccine.

Citation:

Citation:

The sensitivity of ELISA is observed to be much higher compared to the agglutination titers. The ELISA was able to detect specific antibodies at higher serum dilutions at 1:1000 as compared with lower detection limits of the agglutination assay, which could show the positive reaction at 1:256 and 1:64 in bio film and free cell vaccinated fish serum respectively. The results are in agreement with the previous studies of Furuta et al. [15] where they reported the sensitivity of ELISA was six times higher in detecting *Aeromonas hydrophila* than agglutination assay and Yoshimizu et al. [20] reported the sensitivity of ELISA to be several times higher than agglutination assay.

We have shown that the fish vaccinated orally with bio film of *A. hydrophila*, showed elevated antibody titer as well as a good protection against the virulent *A. hydrophila* challenged via intraperitoneal injection. The increased antibody titer and protection against challenged bacteria in bio film vaccinated fish is due to the property of bio film, not being destroyed by the digestive enzymes and being available in large quantity to the lymphoid organs of fish to develop adaptive immune response against the antigen, which is also evident in previous studies [12,13]. Furthermore, the failure to produce enough specific antibodies in free cell vaccinated group is due to the destruction of the antigen by digestive enzymes before reaching the hind gut [12,13]. The immune stimulatory role of chitin, added along with the bio film vaccine may not be denied, as chitin can enhance the innate immune response of the fish [21] which ultimately might have led to the adaptive response to produce increased antibody titer. But, the role of chitin in enhancing the immune response along with bio film needs a detailed study.

With the above results, we conclude that the bio film oral vaccine can be efficiently employed in the culture system to overcome the infectious diseases and ELISA is much more sensitive in detecting specific serum antibodies in vaccinated fish serum. So, it can be used as a tool to evaluate the efficacy of vaccines.

Acknowledgment

The author gratefully acknowledged. Thanks to CIFE, Mumbai, for providing Fellowship to pursue my masters. Help from, Dr Honnananda, Mr. Abhiman and Mr. Ashok (Field assistant) is greatly acknowledged.

References

1. CIFA (2004) Annual Report Central Institute of Freshwater Aquaculture. Bhubaneswar, CIFA.
2. FAO (2005) FISHSTAT Plus – Version 2.30 for 2003 statistics.
3. Ayyappan S & Jena JN (2001) Sustainable freshwater aquaculture in India. In: TJ Pandian (ed) Sustainable Indian Fisheries, National Academy of Agricultural Sciences, New Delhi.
4. Bondad-Reantaso MG, Subasinghe RP, Arthur JR, Ogawa K, Chinabut S, et al. (2005) Disease and health management in Asian aquaculture. *Vet Parasitol* 132: 249-272.

5. Shome R, Shome BR, Mazumder Y, Das A, Kumar A, et al. (2005) Abdominal dropsy disease in major carps of Meghalaya: isolation and characterization of *Aeromonas hydrophila*. *Curr Sci India* 88: 25.
6. Fuda H, Soyano K, Yamazaki F and Hara A (1991) Serum immunoglobulin M (IgM) during early development of masu salmon (*Oncorhynchus masou*). *Comp Biochem Physiol* 99: 637-643.
7. Azad IS, Shankar KM, Mohan CV (1997) Evaluation of an *Aeromonas hydrophila* ELISA for detection of antibody in serum of Indian Major carps. *Dis Aquat Org* 33: 181-186.
8. Azad IS, Shankar KM, Mohan CV, Kalita B (1999) Protective response of Indian Major carps against *Aeromonas hydrophila* challenged by injection and immersion route. *J Aqua Trop* 15: 65-70.
9. Azad IS, Shankar KM, Mohan CV, Kalita B (2000) Uptake and processing of *Aeromonas hydrophila* antigen by Indian Major carps and common carp following of oral vaccination-antigen localization by a monoclonal antibody. *Dis Aquat Org* 43: 103-108.
10. Sureh Babu PP (2008) Characterization of IgM of Indian Major Carps, *Labeo rohita* employing Monoclonal antibody, Doctoral thesis, KVAFSU, Bidar, pp: 80.
11. Furuta T, Iida T, Trongv AK, Sakaguchi J, Akabaashi H (1995) Indirect enzyme linked immunosorbent assay (ELISA) for the detection of antibody in serum of *Aeromonas hydrophila*. *Dis Aquat Org* 16: 447-454.
12. Sanchez C, Fierros PL, Zapata A, Dominguez J (1993) Characterization of monoclonal antibodies against heavy and light chains of trout immunoglobulin. *Dis Aquat Org* 17: 189-194.
13. Rekha M, Shankar KM, Mohan CV (2000) Development of ELISA for detection of carp antibodies. Bangalore.
14. Thuvander A., Hongslo T, Jansson E, Sundquist B (1987) Duration of protective immunity and antibody titers measured by ELISA after vaccination of rainbow trout, *Salmo gairdneri* Richardson, against vibriosis. *J Fish Dis*. 10: 479-486.
15. Yoshimizu M, Direkbusarakom S, Nomura Ezura T, Kimura T (1992) Detection of antibody against *Aeromonas hydrophila* by enzyme linked immunosorbent assay. *Gyobyo Kenkyu* 27: 73-82.
16. Ayyaru G, Venkatesan A (2006) Immunomodulatory effects of dietary intake of chitin, chitosan and levamisole on the immune system of *Cyprinus carpio* and control of *Aeromonas hydrophila* infection in ponds. *Aquaculture* 255: 179-187.